

Branching in pea: double mutants of *rms7* with *rms1* through *rms5*

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Mutants *rms6* and *rms7* in pea have increased branching from basal nodes (6, 10). In contrast, mutants *rms1* through *rms5* have increased branching from both basal and aerial (upper stem) nodes (1-3). A start has been made on checking double-mutant phenotypes. In some cases, the double mutant expresses an additive phenotype with branching more strongly enhanced than in either single mutant, e.g. *rms2-1 rms4-1*, *rms2-1 rms5-2*, *rms3-1 rms6-2* and *rms6-1 rms7-1* (6, 8-10). In other cases, epistasis occurs and the double-mutant phenotype does not transgress beyond the range of the single mutants, e.g. *rms1-1 rms4-1* and *rms2-1 rms3-1* (8).

In the present study, the phenotype of the double mutants of *rms7* with *rms1* through *rms5* was examined in tall (*Le*) plants grown in the glasshouse under an 18-h photoperiod (for details see 8). This strategy was designed to allow identification of double-mutant plants regardless of whether they were clearly obvious from an additive double-mutant phenotype or hidden by epistasis of one or other mutant allele. The strategy makes use of the following information gleaned from years of observation of branching in pea. 1) Basal branching is expressed more strongly in dwarf (*le*) than tall (*Le*) plants (4, 7, 8). 2) In contrast to dwarf plants, tall plants with WT (wild-type) branching genes invariably fail to produce secondary stems from a basal node under the 18-h conditions used. 3) Tall *rms1* through *rms5* plants always produce aerial laterals under these 18-h conditions. Thus in an F₂ population, any tall plant with a major secondary stem and no aerial laterals could be considered as homozygous for *rms7*. In cases where double-mutant plants were not exposed in F₂ by an additive phenotype, F₃ progenies could be grown from the homozygous *rms7* plants and any F₃ plants expressing strong growth of aerial branches would be exposed as double mutants.

In accordance with this strategy, dwarf line M3T-475 (*rms7 1*) was crossed with tall lines Wt15240 (*rms1-5*, ex Kaliski), K524 (*rms2A*, ex Torsdag), K487 (*rms3-1* ex Torsdag), K164 (*rms4-1*, ex Torsdag) and HL298 (*rms5-3*). HL298 was specifically bred for this purpose from a cross between tall cv. Torsdag and Wt15241 (*rms5-3*, ex dwarf cv. Paloma). Further details on these mutant lines are given by Arumingtyas et al.(2).

The *rms1-5 rms7-1* double mutant was found to have an additive phenotype (Fig. 1). Tall F₂ plants of cross Wt15240 (*rms1-5*) x M3T-475 (*rms7-1*) could be partitioned into four branching classes corresponding to WT, *rms7*, *rms1*, and *rms1 rms7* double-

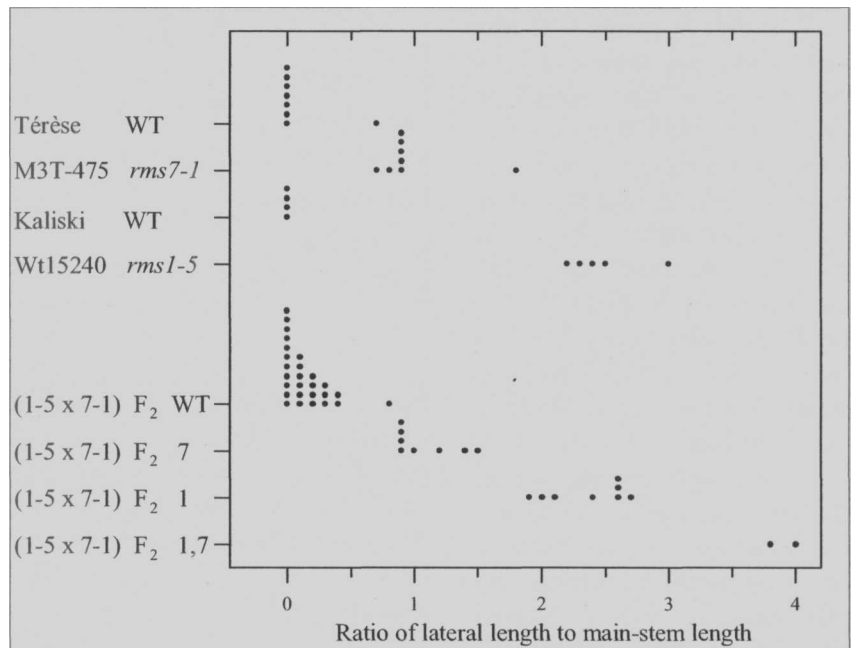


Fig. 1. Distribution of the branching index 'ratio of lateral to main-stem length' for cv. Terese, M3T-475 (*rms7-1*), cv. Kaliski, Wt15240 (*rms1-5*), and tall F₂ plants from the cross Wt15240 x M3T-475. The F₂ data are subdivided into four branching phenotypes representing WT, *rms7*, *rms1*, and double-mutant *rms1 rms7* plants. Data are from mature plants; photoperiod 18 h.

mutant plants. There was a quantum increase in the ratio of lateral to main-stem length from WT to *rms7* to *rms1* to *rms1 rms7*. The observed F₂ numbers of 27 WT, eight *rms7*, eight *rms1* and two *rms1 rms7* plants are in good accordance with a di-hybrid 9: 3: 3: 1 ratio ($P > 0.9$). The additive phenotype of the double mutant was confirmed by growing F₃ progenies from *rms7* F₂ plants (data not shown). The tall *rms1-5 rms7-1* segregates in the F₃ population had a branching index 42 % higher than the WT15240 *rms1-5* single-mutant controls grown with this generation.

Over half the tall WT F₂ plants of cross WT15240 x M3T-475 produced some lateral branches in contrast to the complete absence of branching in the tall WT control Kaliski (Fig.1). The WT F₂ plant with the rather high branching index of 0.8 was confirmed to be WT by growing F₃. The branching in these WT plants was comprised principally of short, late emerging, aerial laterals; no WT plant produced a secondary stem from a basal node. The late outgrowth of aerial laterals on many of the tall WT F₂ plants may partially be explained by the fact that Terese and M3T-475 tended to flower at node 18 while Kaliski and Wt15240 generally flowered at node 16. A two-node delay in flower initiation allowed increased opportunity for aerial lateral outgrowth. (NB. The late emerging aerial laterals referred to here are a normal part of the first reproductive cycle and not to be confused with second-growth laterals that emerge if plants fail to undergo monocarpic senescence).

Growth of laterals from the cotyledonary node (node 0) was rare in the tall F₂ plants of cross Wt15240 x M3T-475 and occurred only in two of the eight *rms7* plants and one of the two double-mutant plants.

The tall F₂ population of cross K524 (*rms2-1*) x M3T-475 (*rms7-1*) gave a clear separation into 21 WT, seven *rms7*, seven *rms2* and two probable *rms2 rms7* plants, numbers that closely fit a 9: 3: 3: 1 ratio (Fig. 2). The two F₂ plants with the high branching indices of 3.8 and 4.4 were backcrossed to K524 and M3T-475; the F₁ results gave supporting evidence these two plants had a double-mutant genotype (data not shown). Interestingly, the double-mutant F₃ plants had a branching index only 8% higher than the K524 *rms2-1* controls grown with them (Fig. 2). Thus there is evidence that the *rms2-1 rms7-1* double mutant has an additive phenotype. However, while the F₂ data indicated a clear quantum increase in branching, the F₃ data showed only a small quantitative increase in branching over the *rms2A* single mutant.

The F₂ of cross K487 (*rms3-1*) x M3T-475 (*rms7-1*) gave no evidence of transgression (data not shown). An *rms3-1 rms7-1* double-mutant line was obtained in F₃ and F₄ via a clear *rms7* F₂ plant. The branching index of the double-mutant plants did not appear to be enhanced beyond the range of vigorous, single-

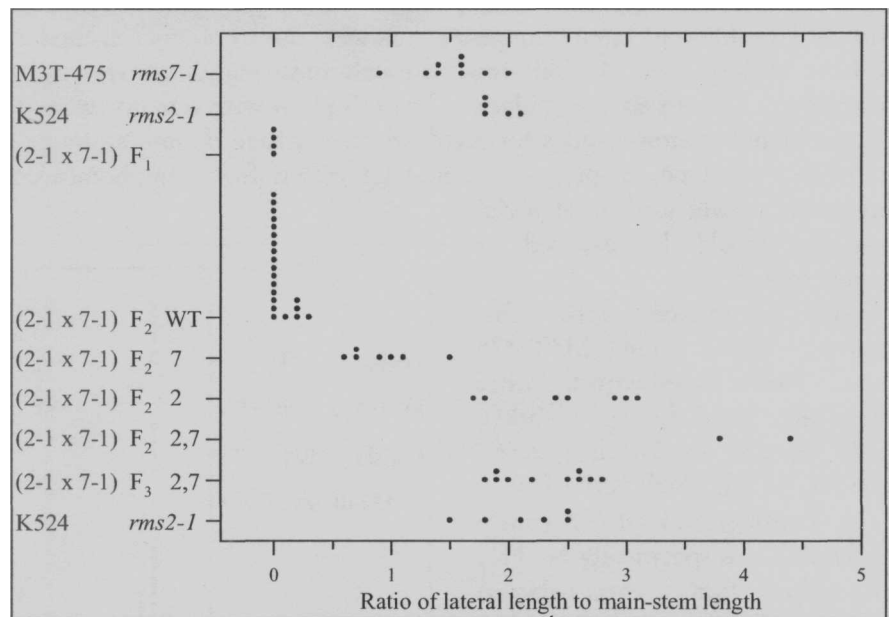


Fig. 2. Distribution of the ratio of lateral to main-stem length for M3T-475 (*rms7-1*), K524 (*rms2-1*), and the F₁, F₂ and F₃ plants of cross K524 x M3T-475. F₂ data are subdivided into four branching phenotypes representing WT, *rms7*, *rms2* and *rms2 rms7* plants. F₂ and F₃ data are from tall plants only. The plants represented in the upper seven rows were sown July 31, 2000 and in the lower two rows July 30, 2001. Data are from mature plants; photoperiod 18 h.

mutant, *rms3-1* plants. However, the double mutant did combine features from both single mutants: the aerial lateral growth of *rms3-1* plants and the tendency to produce laterals from the cotyledonary node shown by *rms7-1* plants. Out of 73 tall *rms3-1 rms7-1* plants, one third produced one or more laterals from node 0, and many of these laterals continued growth into secondary stems. In contrast, basal laterals were not produced from node 0 of the K1487 *rms3-1* mutant: when present, basal laterals grew from nodes 2 and/or 1 of K487 plants.

No transgression for the ratio of lateral to main-stem length occurred in the F₂ of cross K164 (*rms4-1*) x M3T-475 (*rms7-1*) (data not shown). Homozygous *rms4-1 rms7-1* plants were obtained in the F₃ and F₄ via clear *rms7* F₂ plants. The branching index of tall *rms4-1 rms7-1* plants did not transgress beyond the upper range of the K164 *rms4-1* single mutant. Five per cent (3/59) of double-mutant plants produced a lateral from the cotyledonary node, a feature not seen in the single mutant *rms4-1*.

F₂ data for cross HL298 (*rms5-3*) x M3T-475 (*rms7-1*) indicated an additive double-mutant phenotype (Fig. 3). The tall F₂ population could be partitioned into 33 WT, three *rms7*, ten *rms5*, and two probable *rms5 rms7* double-mutant plants with a substantially higher branching index than either single-mutant parent. The phenotypic contrast between the *rms5* single mutant and the double mutant is not wholly revealed by the index 'ratio of lateral to main-stem length'. Expression of *rms5-3* in a tall background seems fairly weak. Three out of six HL298 plants produced no basal lateral of any consequence; likewise three out of ten tall *rms5* F₂ plants. These *rms5* plants were not really identifiable as *ramosus* mutants until outgrowth of aerial laterals commenced three to four weeks after sowing. In contrast, the presumed double-mutant plant with index 3.5 (Fig. 3) had four basal laterals exceeding 10 mm by day 11 (two from node 0, one from node 1 and one from node 2) and all four shoots continued growth to become secondary stems. These observations provide further support for the view that *rms5-3 rms7-1* plants have an additive phenotype that combines the aerial branching of *rms5-3* plants with the enhanced basal branching of *rms7-1* plants.

Segregation for branching phenotype in the F₂ of cross HL298 x M3T-475 (Fig. 3) is in accordance with a 9: 3: 3: 1 ratio ($P > 0.1$). However, numbers in the *rms7* class are below expectation and it seems highly likely that some tall *rms7* plants in this F₂ failed to produce a basal lateral and were indistinguishable from WT plants. In a tall background, results from several crosses showed that *rms7-1* behaved as a weak mutant lacking the full penetrance of classic Mendelian mutants. In the F₃ of cross Wt15240 (*rms1-5*) x M3T-475 (*rms7-1*), only 38% (8/21) of tall *rms7* plants had a phenotype distinguishable from WT. In the F₃ of crosses K164 (*rms4-1*) x M3T-475 and K487 (*rms3-1*) x M3T-475, penetrance of *rms7-1* fell to 15% (3/20) and 6% (2/36), respectively. With a penetrance that low, the *rms7-1* mutation may well have escaped detection had it been induced in a tall cultivar. We were also fortunate to have obtained *rms7* numbers right on Mendelian expectation in two F₂ populations (Figs 1 and 2). Clearly, penetrance of *rms7-1* varied from planting to planting.

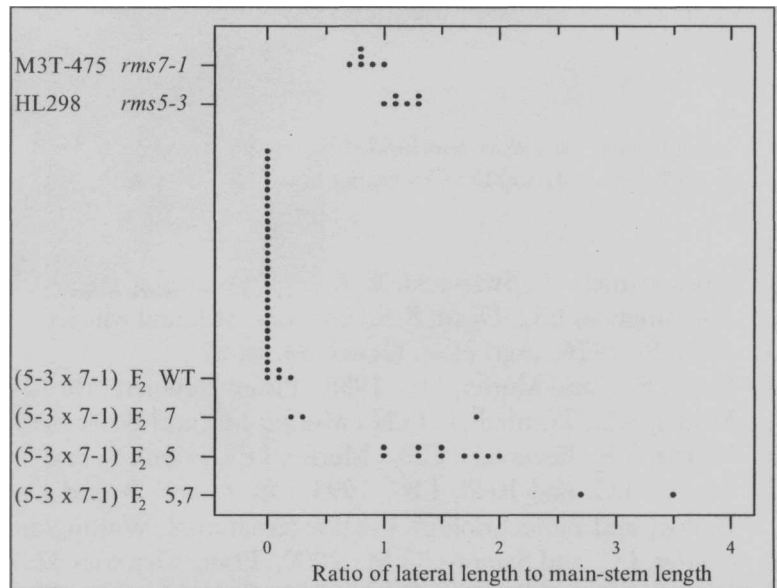


Fig. 3. Distribution of the ratio of lateral to main-stem length for M3T-475 (*rms7-1*) and HL298 (*rms5-3*), and tall F₂ plants from the cross HL298 x M3T-475. The F₂ data are subdivided into four branching phenotypes representing WT, *rms7*, *rms5*, and double mutant *rms5 rms7* plants. Data are from mature plants; photoperiod 18 h.

In summary, a clearly additive double-mutant phenotype was observed for *rms1-5 rms7-1* (Fig. 1) and *rms5-3 rms7-1* (Fig. 3), *rms2-1 rms7-1* showed variable levels of transgression (Fig. 2), and *rms3-1 rms7-1* and *rms4-1 rms7-1* did not transgress, respectively, beyond the upper range of the *rms3-1* and *rms4-1* single-mutant parents. Interestingly, *rms1* and *rms5* both produced an additive phenotype with *rms7* (Figs 1 and 3), which fits well with evidence that *RMS1* and *RMS5* regulate the same novel branching signal (5). However, *rms1-5* and *rms5-3* had a lower branching index than *rms2-1*, *rms3-1* and *rms4-1* when all five mutants were planted and grown together (data not shown). Thus *rms1-5* and *rms5-3* have more room to show enhanced branching and an additive phenotype in combination with *rms7-1*.

The basal branching mutants *rms6-1* and *rms7-1* produced an additive double-mutant phenotype, suggesting that *RMS6* and *RMS7* may operate in different pathways (6). That view is supported by the fact that *rms3-1 rms6-2* was found to have an additive phenotype with strongly enhanced branching (10), whereas no evidence of transgression was found here for the *rms3-1 rms7-1* double mutant.

Acknowledgements: This work was funded by a grant from the Australian Research Council. I thank Dr Suzanne Morris for help with Figs. 1-3, and Ian Cummings and Tracey Winterbottom for technical assistance.

1. Apisitwanich, S., Swiecicki, W.K. and Wolko, B. 1992. *Pisum Genetics* 24: 14-15.
2. Arumingtyas, E.L., Floyd, R.S., Gregory, M.J. and Murfet, I.C. 1992. *Pisum Genetics* 24: 17-31.
3. Blixt, S. 1976. *Agri Hort. Genet.* 34: 83-87.
4. Floyd, R.S. and Murfet, I.C. 1986. *Pisum Newslett.* 18: 12-15.
5. Morris, S.E., Turnbull, C.G.N., Murfet, I.C. and Beveridge, C.A. 2001. *Plant Physiol.* 126: 1205-1213.
6. Morris, S.E., Beveridge, C.A., Murfet, I.C., Prioul, S. and Rameau, C. 2003. *Pisum Genetics* 35: 10-14.
7. Murfet, I.C. and Reid, J.B. 1993. In: Casey, R and Davies, D.R. (eds.) *Peas: Genetics, Molecular Biology and Biotechnology*. CAB International, Wallingford, UK, pp 165-216.
8. Murfet, I.C. and Symons, G.M. 2000. *Pisum Genetics* 32: 33-38.
9. Murfet, I.C. and Symons, G.M. 2000. *Pisum Genetics* 32: 59-60.
10. Rameau, C, Murfet, I.C, Laucou, V., Floyd, R.S., Morris, S.E. and Beveridge, C.A. 2002. *Physiol. Plant.* 115:458-467.