

Mapping of the new mutation *blb* and the problem of integrity of linkage group I

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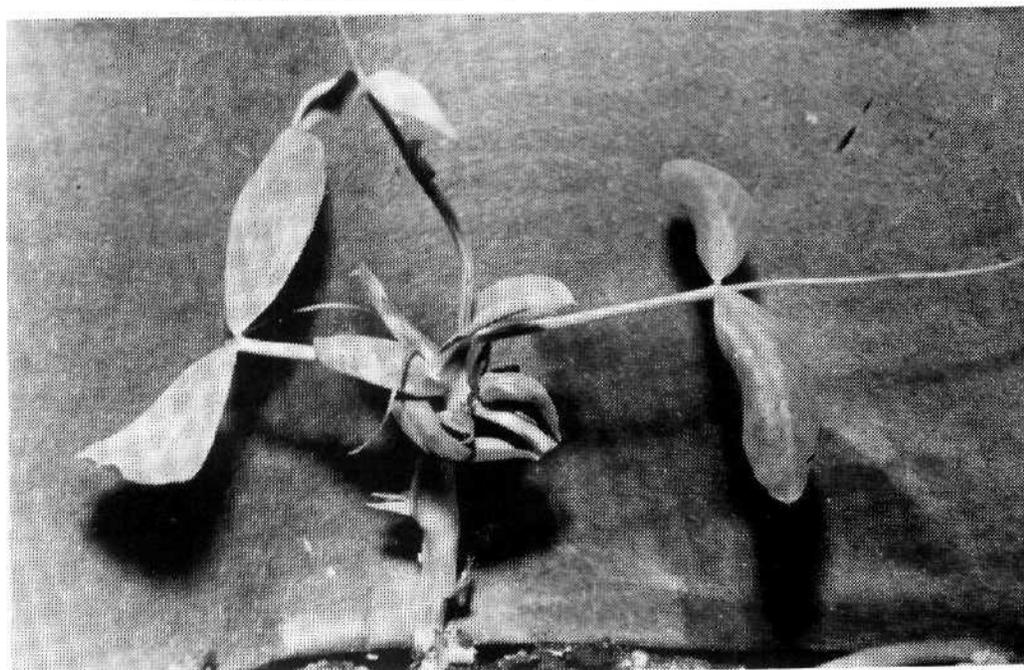
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A new recessive mutant, designated *blb* (*bulbosus*), is described and mapped on linkage group I.

The *blb* mutation was induced in line SGE following treatment with 0.15% EMS (ethylmethanesulphonate). The original mutant line was registered as SGE80. The initial line SGE was derived by V.A. Berdnikov from crosses among lines VIR6135 (from Greece), VIR320 (from Palestine, *Pisum sativum syriacum*), and an original ultra rapid line, Sprint-1.

The *blb* mutation transforms the shape of leaflets and stipules. The leaflets of homozygous *blb* plants are rather narrow, especially at lower nodes, and are wedge-shaped so that the maximal width of a leaflet is attained distally from the middle of its length (Fig. 1). At the first several nodes, the leaflet edges are rolled inside. The stipules are narrowed and, as a rule, do not overlap each other. The pods are narrowed, especially in the base, and seed productivity is reduced. In seedlings of the original mutant line SGE80 the stem is slightly thickened just above the first scale leaf. However, in the F₂ progenies of three crosses involving this mutant (see below) this feature was much more pronounced. The seedlings homozygous for *blb* appeared to be considerably delayed in their growth, and their stems within the first 2-3 nodes were shortened and extremely inflated - up to 1 cm wide (Fig. 1). The name *bulbosus* was given to the mutant due to this unusual swelling of the stem. It was our impression that the apex ceased to grow and all the nutrients were directed into stem inflation. In the F₂ progeny of cross SGE80 x WL1238, which was grown in a greenhouse, the apical buds of homozygous *blb* plants usually died and several lateral branches grew from basal nodes. In the F₂ of cross VIR3971 x SGE80, which was planted in the field, mutant seedlings ceased growth for several days and then produced a normal shoot from the apical bud. In this cross the mutant plants usually produced no basal branches, while their counterparts with a normal phenotype exhibited vigorous basal branching. The situation was therefore opposite to that observed in the previous cross. The degree of stem inflation in mutant seedlings was variable and some *blb* homozygotes had stems only slightly thickened, as in the original mutant line SGE80. The modified leaflet shape was, however, an invariable character of *blb* homozygotes.

The F₂ plants of cross SGE80 x WL1238 were tested for histone H1 electrophoretic spectrum, as described in (1, 2). The parental lines have different allelic variants of H1 subtype 7: SGE80 has allele *His7*² and WL1238 has allele *His7*³. A significant linkage of 20 cM was detected between loci *blb* and *His7* (Table 1) suggesting that *blb* belongs to linkage group I. Unfortunately, no other neighbouring marker genes were involved in this cross.



Figs 1a and b. Seedlings homozygous for *blb*.

Table 1. Joint segregation of genes *His7* and *blb* in the F₂ of cross SGE80 x WL1238.

	<i>His7</i> ³	<i>His7</i> ³ / <i>His7</i> ²	<i>His7</i> ²
<i>Blb</i>	15	37	7
<i>blb</i>	1	5	11

Recomb. Fract = 19.91%; SE = 5.04%; Linkage $\chi^2 = 20.60$; P = 0.00003.

In order to map gene *blb*, two other crosses were carried out: VIR3971 x SGE80 and SGE80 x OK7. Line SGE80 has alleles *blb*, *His(2-6)*¹²²¹ [this is the cluster of genes encoding histone H1 subtypes 2-6 (1, 2)], *His7*², *A*, *D*^w, and *i*; line VIR3971 has alleles *Blb*, *His(2-6)*¹¹¹¹, *His7*¹, *A*, *D*^{co}, and *I*; line OK7 (derived by O. Kosterin from Weibullsholm lines WL1393, WL1688, and WL102) has alleles *Blb*, *His(2-6)*¹¹²¹, *His7*³, *a*, *D*^{co}, and *I*. In both crosses, the F₁ hybrids were vigorous and exhibited full fertility of both pollen and ovules. We grew F₂ progenies comprising 663 plants of the former cross and 123 plants of the latter. F₂ segregation data are presented in Tables 2 and 3.

The recombination frequencies inferred from the cross VIR3971 x SGE80 indicate the following arrangement of genes:

$$\begin{array}{ccccccc}
 \textit{His}(2-6) & \text{-----} & \textit{His}7 & \text{-----} & \textit{blb} & & \\
 & \text{---} 28.1 \pm 1.5 \text{---} & & \text{---} 17.6 \pm 1.6 \text{---} & & & \\
 & & & \text{-----} 36.6 \pm 2.2 \text{-----} & & & \text{cM}
 \end{array}$$

The distance between *His(2-6)* and *His7* corresponds well to that obtained in several other crosses (2, 3, and unpub.).

The data from cross SGE80 x OK7 indicate the following map segment:

$$\begin{array}{ccccccc}
 \textit{His}(2-6) & \text{---} \textit{a} & \text{-----} & \textit{His}7 & \text{-----} & \textit{blb} & \\
 & \text{---} 13.6 \pm 3.3 \text{---} & & \text{---} 33.0 \pm 4.0 \text{---} & & \text{---} 18.7 \pm 3.8 \text{---} & \\
 & & & \text{-----} 35.6 \pm 4.0 \text{-----} & & & \\
 & & & & \text{-----} 33.8 \pm 7.9 \text{-----} & & \\
 & & & \text{-----} 36.7 \pm 5.2 \text{-----} & & & \text{cM}
 \end{array}$$

This map shows a good correspondence to the previous one, but, for an unknown reason, the distance *His(2-6)* – *a* is much greater than the 5-7 cM observed in numerous other crosses (1, 2, and unpub.).

Table 2. Joint segregation data obtained from the F₂ of cross VIR 3971 x SGE80.

Gene A	Gene B	Phenotype ¹									Recomb. Fract. (%) ²	SE
		A/B	A/h	A/b	h/B	h/h	h/b	a/B	a/h	a/b		
<i>His</i> (2-6)	<i>His7</i>	97	63	11	81	188	52	23	59	89	28.12	1.52***
<i>blb</i>	<i>His7</i>	198	268	56				3	42	96	17.63	1.61***
<i>blb</i>	<i>His</i> (2-6)	154	256	112				17	65	59	36.59	2.23***
<i>blb</i>	<i>d</i>	399		123	(repulsion)			109		32	49.60	2.93
<i>blb</i>	<i>i</i>	346		176	(coupling)			95		46	51.80	2.97
<i>d</i>	<i>His</i> (2-6)	125	248	135				46	73	36	53.07	2.37
<i>d</i>	<i>His7</i>	118	227	163				34	83	38	52.34	2.37
<i>d</i>	<i>i</i>	342		166	(repulsion)			99		56	51.61	2.86
<i>i</i>	<i>His</i> (2-6)	103	224	114				68	97	57	52.74	2.37
<i>i</i>	<i>His7</i>	125	209	107				76	101	45	54.43	2.36

¹ In the case of dominant genes, the letters A and B stand for the dominant alleles and the letters a and b for the recessive ones. Where the second gene is codominant, capital A stands for the dominant allele of the first gene and capital B for an allele of the second gene which is in coupling with A. Where both genes are codominant, the capital letter stands for an allele coming from the first parent, h stands for heterozygotes.

² The recombination fractions and their standard errors were estimated by Fisher's method of maximum likelihood. In this and the following table ** and *** indicate that the recombination values differ from 50% by more than two and three times the standard errors, respectively.

Note: the plants also segregated for genes *fl* (group 6) and *His1* (group 5); these genes segregated independently of each other and of the other genes involved.

Table 3. Joint segregation data obtained from the F₂ of cross SGE80 x OK7.

Gene A	Gene B	Phenotype ¹									Recomb. Fract. (%) ¹	SE
		A/B	A/h	A/b	h/B	h/h	h/b	a/B	a/h	a/b		
<i>a</i>	<i>His</i> (2-6)	28	47	8				1	8	31	13.65	3.29***
<i>His7</i>	<i>His</i> (2-6)	6	15	4	21	27	14	2	13	21	35.64	4.02***
<i>blb</i>	<i>His</i> (2-6)	36	42	20				3	13	9	36.69	5.17**
<i>a</i>	<i>His7</i>	21	45	17				4	17	19	32.96	4.98***
<i>a</i>	<i>blb</i>	62		21	(repulsion)			36		4	33.79	7.87**
<i>blb</i>	<i>His7</i>	36	52	10				0	10	15	18.68	3.84***
<i>blb</i>	<i>d</i> ²	44		17	(repulsion)			17		4	43.04	8.90
<i>d</i> ²	<i>His7</i>	17	31	13				4	14	3	49.39	6.76
<i>d</i> ²	<i>His</i> (2-6)	22	34	5				6	12	3	44.57	6.69

¹ See footnotes for Table 2.

² Joint segregations involving *d* were analysed only among A plants.

Earlier (3) we demonstrated the absence of linkage between genes *His7* and *d*. Linkage between these genes should be detected if the published maps of pea linkage group I are correct (the expected distance varies from 1-20 cM depending on the particular map version). Moreover, we found (3) that placement of genes *a* and *d* in the same linkage group was based on ambiguous data concerning their direct segregation and very questionable data concerning segregation of gene *au*. The segregation data from crosses (Tables 2 and 3) show no evidence or indication of linkage between *d* and either *His(2-6)*, *His7*, or *blb*. This result extends by some 17-19 cM the length of the map region in the upper section of group I where linkage with *d* cannot be detected. Likewise, there was no indication of linkage between either *His(2-6)*, *His7*, *blb* or *d* and standard marker *i* which is placed in the lower section of group I in existing linkage maps. Thus our present data further support the previous conclusion of Kosterin (2, 3) that the *d* - *i* linkage segment is not on the same chromosome as the *a*—*His7* linkage group. Gene *blb* now appears to be the most distal (in the direction of gene *lf* from gene *a*) marker among the so far known loci of group I. There is, therefore, an urgent need to obtain additional linkage data on the *d* - *i* segment of the pea genome.

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 2. Kosterin, O.E. 1992. *Pisum Genetics* 24:56-59.
 3. Kosterin, O.E. 1993. *Pisum Genetics* 25:23-26.